

Alfonso J. Chay-Canul¹ 0000-0003-4412-4972Jorge Tapia-González¹Jorge Rodolfo Canul-Solís² 0000-0001-9934-4302Fernando Casanova-Lugo³ 0000-0003-2485-9170Ángel T. Piñeiro-Vázquez⁴ 0000-0002-8400-4046Rodrigo Portillo-Salgado⁵ 0000-0001-7253-3752Ricardo García-Herrera^{1*} 0000-0003-2456-4727Einar Vargas-Bello-Pérez^{6, 7*} 0000-0001-7105-5752

¹Universidad Juárez Autónoma de Tabasco.
División Académica de Ciencias Agropecuarias.
Villahermosa, Tabasco, México.

²Tecnológico Nacional de México.
Instituto Tecnológico de Tizimín.
Tizimín, Yucatán, México.

³Tecnológico Nacional de México.
Instituto Tecnológico de la Zona Maya. Othón P.
Blanco, Quintana Roo, México.

⁴Tecnológico Nacional de México.
Instituto Tecnológico de Conkal.
Conkal, Yucatán, México.

⁵Colegio de Postgraduados.
Programa en Recursos Genéticos
y Productividad-Ganadería.
Texcoco, Estado de México, México.

⁶School of Agriculture.
Policy and Development New Agriculture Building.
Earley Gate Whiteknights Road, Berkshire,
United Kingdom.

⁷Universidad Autónoma de Chihuahua.
Facultad de Zootecnia y Ecología.
Chihuahua, México.

***Corresponding authors:**

Email address:

ricardo.garcia@ujat.mxevargasb@uach.mx

Predictive biometrics of hair sheep through digital imaging

Abstract

Direct collection of biometric measurements (BM) from sheep is an expensive and stressful procedure for animals; instead, indirect and novel methods have recently been used. The objective of this study was to use digital image analysis (DIA) to predict biometric measurements of Pelibuey sheep as a non-invasive approach under on-farm conditions. Withers height (WH), body length (BL), body diagonal length (BDL), and rib depth (RD) were predicted in Pelibuey ewes using DIA. Images were taken from the left flank of 65 non-pregnant and nonlactating Pelibuey ewes using a digital camera and analyzed by DIA. The BM determined from both *in vivo* and by DIA presented positive and moderate ($P < 0.05$) correlation coefficients (r) of 0.43, 0.66, 0.73, and 0.75 for BL, BDL, WH, and RD, respectively. Regression equations from BM by DIA had determination coefficients (r^2) of 0.19, 0.44, 0.54, and 0.56 for BL, BDL, WH, and RD, respectively. The equations developed were from low to moderate precision ($r^2 = 0.18$ to 0.55), moderate to high accuracy with a bias correction factor ($C_b > 0.69$), and low to moderate reproducibility index (> 0.30). Overall, the use of DIA was able to predict the BM in Pelibuey ewes with low to moderate precision and accuracy. Factors affecting the accuracy and precision of this relationship should be further investigated.

Keywords: Body measurements; Image analysis; Linear regression equations; Image-processing; Body weight; Tropical conditions..

Submitted: 2022-11-30

Accepted: 2023-07-27

Published: 2023-09-06

Additional information and declarations
can be found on page 10

© Copyright 2023
Alfonso J. Chay-Canul *et al.*

open access 

Distributed under Creative Commons CC-BY 4.0

Cite this as:

Chay-Canul AJ, Tapia-González J, Canul-Solís JR, Casanova-Lugo F, Piñeiro-Vázquez ÁT, Portillo-Salgado R, García-Herrera R, Vargas-Bello-Pérez E. Predictive biometrics of hair sheep through digital imaging. *Veterinaria México OA.* 2023;10. doi: 10.22201/fmvz.24486760e.2023.1150.

Study contribution

This study describes an alternative, low-cost method that allows farmers to make decisions when selecting animals based on body weight.

Introduction

Image-processing technologies have developed rapidly and can be used to quantitatively characterize the size, shape, and density of organisms or objects.⁽¹⁾ Currently, digital image analysis (DIA) is used in many fields, such as human medicine,⁽²⁾ veterinary medicine⁽³⁾ and forensic sciences.⁽⁴⁾ Similarly, currently this technology is being used in animal science for carcass characteristics and composition predictions of meat products.^(5, 6) Digital image analysis has been used to estimate body condition, live weight, and carcass composition in heifers,⁽⁷⁾ beef and dairy cattle,⁽⁸⁻¹⁴⁾ sheep,⁽¹⁵⁾ pigs,⁽¹⁶⁾ yaks,⁽¹⁷⁾ camels,⁽¹⁸⁾ and broiler chickens.^(19, 20)

The determination of body weight (BW) is one of the more accurate methods to determine growth and plays an important role in management decisions to support livestock production.^(7, 21) Determination of BW in livestock through image analysis is an emerging research area, allowing the possibility to automatically measure the dimensions of animal images and use prediction equations to establish the relationship between them and live BW.⁽²²⁾ The most common approach is the use of images based on the lateral and top areas of the animal that provides different body measurements such as withers height, rump height, body length, diagonal body length, rid depth, chest depth, chest width, thorax width, abdomen width, and dorsal height, that can be correlated with BW.^(8, 22, 23)

Various techniques have been reported to measure or estimate the BW of livestock as alternatives to the use of weight scales, which is still the most accurate method used by small farms, but it is time-consuming. Also, this is a stressful management situation that in some cases leads to temporal BW losses from animals, and this management includes large fasting periods and deprivation of water derived from holding the sheep in handling pens and these losses can be from 1.8 to 2.9 kg or from 3.5 to 5.6 % BW.^(21, 24)

Therefore, it is important to develop alternative and practical methods that are low-cost and easy to implement for decision support systems. Among the alternative methods, the use of biometric measurements (BM) has previously been suggested.⁽²⁴⁻²⁶⁾ Some authors have reported that manual measurement of BM or body condition score (BCS) of animals is a time-consuming and stressful task for both the farmer and the animal.^(7, 27) Recently the use of DIA has been applied to determine the BM and BW of cattle.^(8, 17, 28-30) However, in sheep breeds, the use of DIA to predict their BM, BW, and body size is limited and has not been used in on-farm situations.⁽¹⁵⁾ Moreover, it has been reported that in tropical conditions, sheep production systems are characterized by low inputs, and most hair sheep breeds are used. Compared with wool breeds, hair sheep breeds such as Pelibuey are small, with a slow growth rate and poor muscular conformation.⁽³¹⁾ The objective of this study was to use DIA to predict biometric measurements in Pelibuey sheep as a non-invasive approach under on-farm conditions.

Materials and methods

Ethical statement

Animals were handled according to the guidelines and regulations for animal experimentation of the Department of Agricultural and Livestock Sciences (División Académica de Ciencias Agropecuarias) of the Autonomous Juárez University of Tabasco (Universidad Juárez Autónoma de Tabasco).

Animals, diets, and handling

The data and digital images were collected from 65 nonpregnant and nonlactating Pelibuey ewes aged 2 to 4 years. Animals were selected from a flock composed of 407 adult sheep of the "El Rodeo" farm located at 17° 84' N latitude and 92° 81' W longitude, km 14 along the Villahermosa-Jalapa highway in the "Rancheria Víctor Manuel Fernández Manero" in Jalapa, Tabasco, México. The sheep were grouped in pens within a roofed building with a concrete floor and no walls. The diet consisted of a mixed ration of 66 % forage and 34 % concentrate, with an estimated metabolizable energy of 0012 MJ/kg dry matter (DM) and 10 % crude protein (CP)⁽³²⁾. The dietary ingredients were cereal grains (corn or sorghum), soybean meal, and hay of tropical grasses, vitamins, and minerals.

Biometric measurement evaluation

Individual biometric measurements were taken before feeding at 08:00, considering the BM described by Bautista-Díaz et al.⁽²⁵⁾ For that, withers height (WH) was measured from the highest point over the scapulae, vertically to the ground. Body length (BL) was measured as the distance between the dorsal point of the scapulae and the ventral point of the tuber coxae. Body diagonal length (BDL) was measured as the distance between the ventral point of the tuber coxae and the cranial point of the shoulder, and finally, rib depth (RD) was measured vertically from the highest point over the scapulae to the endpoint of the rib (at the sternum) (Figure 1). All BMs were recorded in cm. Flexible fiberglass tape and a 65 cm caliper were used for measurements. In addition, the animals were weighed using a digital scale (EQB Model, Torrey, México). Body weight and BM were taken from the animals while standing in a sheep chute.

Digital image analysis

A digital camera with 10.2 megapixels (Sony DSLR-A200, San Diego, USA) was used to take photographs. The camera was set to a standard quality (3872 4000 by 2 592 pixels resolution). The images were taken immediately after the BM was recorded on the same day for all animals. The distance between the digital camera and the chute was the same for all animals, and a metal ruler of 30 cm was fixed on the sheep chute and was used for calibration as described by Ozkaya⁽³⁰⁾ (Figure 1). Three photographs were taken of each animal from the left flank. The best image was selected using the criteria that the entire body of the ewe should stand inside the rectangular area set up and that the image should be of good (clear and steady) resolution as described by Wongsriworaphon et al.⁽³³⁾ (Figure 1).

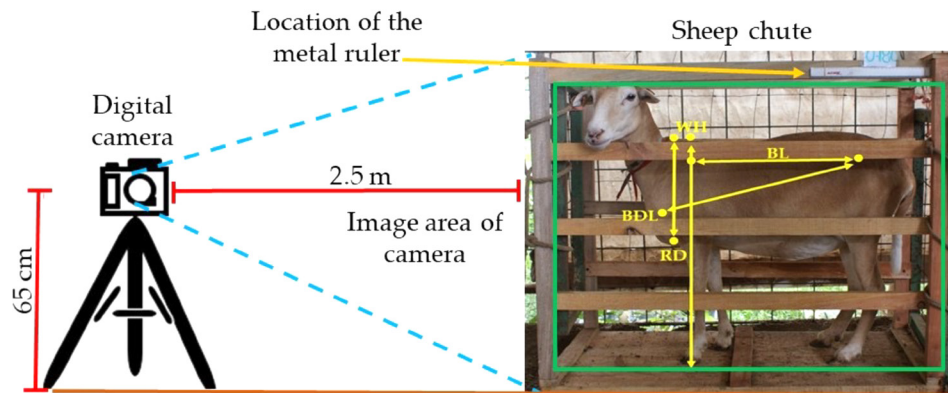


Figure 1. Digital camera position (length and distance from the target animal) for biometric measurements from hair sheep. WH: withers height, BL: body length, BDL: body diagonal length, and RD: rib depth. The green rectangle is the area where the animal should stand for image capture.

The images were manually measured using ImageJ software ver. 1.51 (<https://imagej.nih.gov/ij/index.html>) to determine BM calibrated in centimeters. The linear parameters of the lateral profile of WH, BL, BDL, and RD were measured using the tools of the software to DIA as described by Ozkaya.⁽³⁰⁾ The same operator performed the image processing. To orient his shot optimally, the operator placed a mobile cursor at the two ends of each measuring BM. The length of the line was scaled automatically from a pre-programming setup using a 30 cm-long metal ruler for each image.

Statistical Analysis

Descriptive statistical analysis was performed using the MEANS procedure of SAS ver.9.3. Correlation coefficients (r) between variables were estimated using the SAS CORR procedure. The relationships between BM determined by DIA and BM determined *in vivo* were estimated with linear regression models using the SAS GLM procedure. The precision was assessed by evaluating the r^2 of the linear regression of Y (i.e., observed) on X (i.e., predicted).

Also, to assess the predictability of the equations, coefficients of determination (r^2), mean square error (MSE), standard deviation (SD), mean square error of prediction (MSEP), and root of the MSEP (RMSEP) were performed and accounted for the distance between predicted values and true values.⁽³⁴⁾ The mean bias (MB), as described by Cochran and Cox,⁽³⁵⁾ was used as a representation of the average inaccuracy of the model, and its calculation is based on the mean difference between observed and model-predicted values.⁽³⁵⁾

The modeling efficiency factor (MEF), which represents the proportion of variation explained by the line $Y = X$, was used as an indicator of goodness of fit.^(36,37) The coefficient of model determination (CD) is the ratio of the total variance of observed data to the squared difference between the model-predicted and mean of the observed data. Therefore, the CD statistic explains the proportion of the total variance of the observed values explained by the predicted data.⁽³⁴⁾ The closer to unity the better the model predictions ($CD > 1$ indicates under prediction and $CD < 1$ indicates over prediction).

Table 1. Descriptive statistics of biometric measurements of Pelibuey sheep were determined *in vivo* and by digital image analysis (n = 55)

Variable	Description	Mean	SD	CV	Maximum	Minimum
Observed measurements						
WH	Observed withers height	63.81	3.98	6.23	70.00	56.00
BL	Observed body length	48.67	8.01	16.47	55.00	46.00
BDL	Observed body diagonal length	59.60	9.29	15.59	68.00	56.00
RD	Observed rib depth	34.28	5.36	15.63	42.00	31.00
DIA measurements						
WH _{DIA}	Predicted withers height	59.05	3.48	5.90	67.65	48.39
BL _{DIA}	Predicted body length	37.84	3.37	8.90	45.47	31.81
BDL _{DIA}	Predicted body diagonal length	54.21	4.53	8.36	65.81	44.95
RD _{DIA}	Predicted rib depth	29.64	2.52	8.49	35.20	24.96

SD: standard deviation; CV: coefficient of variation; DIA: digital image analysis

Bias correction factor (Cb), a component of the concordance correlation coefficient (CCC),⁽³⁸⁾ was used as an indicator of deviation from the identity line, and the CCCs were also used as a reproducibility index to account for accuracy and precision. High accuracy and precision were assumed when the coefficients were > 0.80, and low accuracy and precision when the coefficients were < 0.50. Finally, all calculations were obtained using the Model Evaluation System.⁽³⁴⁾

Results and discussion

To our knowledge, the present study is the first to evaluate equations for predicting biometric measurements of Pelibuey sheep as a non-invasive approach under tropical on-farm conditions. After image analysis, data from 10 animals were removed because their images had a low resolution for their correct analysis. This was because when the image was taken, animals were not borne equally on all four feet; they were turned or kneeled. Mean values, SD, and coefficient of variation (CV) of the BM determined *in vivo* and by DIA are presented in Table 1. The measurements determined *in vivo* and by DIA were positive and moderately ($P < 0.0500$) correlated with values of 0.43, 0.66, 0.73, and 0.75 for BL, BDL, WH, and RD, respectively. The digital body measures were highly correlated (> 0.95) between each image labeling.

The regression equations to predict the BM determined *in vivo* using those obtained by DIA had an r^2 of 0.19 for BL, 0.44 for BDL, 0.54 for WH, and 0.56 for RD (Table 2). Additionally, we explored the relationships between BW and BM obtained by DIA but they were not significant. The results of the predictive equations developed in the present study based on the coefficient of determination, suggest that the use of digital image processing had moderate potential to estimate biometric measurements in hair sheep. Compared with the results of Ozkaya⁽³⁰⁾ who reported that the DIA provided very close measurements and therefore highly related to those determined *in vivo* in female Holstein calves ($r^2 > 0.99$), our results were quite different.

For the evaluation of the developed equations (Table 3; Figure 2), the equations had low to moderate precision (; Table 3). However, all equations had moderate to high accuracy (bias correction factor > 0.69; Table 3; Figure 2), confirming a moderate reproducibility index and concordance with the observed data (CCC = > 0.60) except the equation for BL (CCC = 0.30). In addition, the positive mean bias (difference between observed and model-predicted values) and the CD value (CD > 1) indicates an under-prediction, suggesting a shift of the model-predicted values to the left of the $Y = X$ line.⁽³⁴⁾ Relating to the MEF, the equations presented a low efficiency of prediction (from 0.13 to 0.55) and indicated a low concordance between the observed and predicted values considering that a perfect fit is equal to 1.

The CD ranged from 1.79 to 4.22, indicating high variability in the predicted data. In this case, a CD value > 1 indicates under-prediction and a CD value < 1 indicates over-prediction (perfect fit = 1).⁽³⁴⁾ In all equations, a small proportion of the error of prediction was associated with the slope, and the main component of MSEP was a random error (> 94.08 %), which indicated small biases in the predictions. In all equations, the RMSEP accounted for 3.78 % to 4.93 % of the observed BM. Moreover, the null hypothesis (intercept = 0 and slope = 1) was accepted for all equations. The prediction model for BL presented the lowest precision ($r^2 = 0.19$), which according to Na Zhang et al.,⁽¹⁸⁾ could be due to the change in the animal's body posture at the time of shooting the photographs. This factor has a great influence on body length and therefore, these authors recommend taking at least five photographs at different times and positions.

In this study, results from the regression equations that were produced had poor predictive capacity. That may be related to potential errors such as: a) an operator error, b) uncertainty in cursor placement during DIA, c) scaling error because of the scale being at one distance and focal length, and the points of measurement being a further distance and the impact of perspective, and d) animal body position between the *in vivo* measurements and DIA image capture that might lead to different answers from repeated measurements and different postures. In this regard, Khojastehkey et al.⁽¹⁸⁾ indicated that the precision of the model is affected by various factors, such as image quality, the type of statistical model, the number of records, the nature of the biometric measurement, and the position and posture of animals. This last factor has been reported in a similar study where live weight in pigs was estimated based on DIA.⁽³³⁾ Also, some authors noted that the quality of the prediction of live weight is somewhat sensitive to the environment and the animal's position^(33,39) and stated that in field conditions these requirements are not practical and difficult to obtain. The dilemma in this case is that under on-farm conditions, a controlled environment will be difficult but some of the other variables may be easier to modulate.

To improve precision and accuracy, some technical issues need to be considered. Lasfeto and DaudLetik⁽⁴⁰⁾ reported that to obtain the right animal body dimensions and body weight, it is fundamental to perform quality checks and calibration of cameras used for the analysis. The pixel values of an animal's image greatly affect the actual values for body weight and body length. On the other hand, the small sample size used in the present study also influenced the poor predictive capacity of the models. Previously, Gomes et al.⁽⁸⁾ demonstrated that the models to estimate body weight in beef cattle through DIA could have better precision using a greater number of animals ($r^2 = 0.84$ for 35 animals vs $r^2 = 0.69$ for 15 animals).

Table 2. Regression equations to predict biometric measurements of Pelibuey sheep using digital image analysis

	Equations	MSE	RSD	r ²	P-value
1	$WH (cm) = 14.43(\pm 6.37^*) + 0.84(\pm 0.11^{***}) \times WH_{DIA}$	7.46	2.73	0.54	< 0.0001
2	$BL (cm) = 38.85(\pm 3.70^{***}) + 0.29(\pm 0.09^{**}) \times BL_{DIA}$	4.32	2.08	0.19	0.0050
3	$BDL (cm) = 36.70(\pm 4.07^{**}) + 0.44(\pm 0.07^{**}) \times BDL_{DIA}$	5.35	2.31	0.44	< 0.0001
4	$RD (cm) = 12.22(\pm 2.80^{**}) + 0.76(\pm 0.09^{**}) \times RD_{DIA}$	3.00	1.73	0.56	< 0.0001

P-value: *P < 0.0500, **P < 0.0100, ***P < 0.001. r²: determination coefficient to predict the biometric measurement; MSE: mean square error; RSD: residual standard deviation. Values in parentheses are the standard errors (SEs) of the parameter estimates. WH: withers height, BL: body length, BDL: body diagonal length and RD: rib depth; DIA: Digital Image Analysis

Table 3. Mean and descriptive statistics of the accuracy and precision of the equations for predicting biometric measurements of Pelibuey sheep using digital image analysis (n = 55)

Variable ¹	WH	BL	BDL	RD
Mean	63.4	49.3	60.5	34.6
Standard deviation	2.89	0.97	2.00	1.91
Maximum	70.5	51.5	65.6	38.9
Minimum	54.5	47.6	56.5	34.6
r ²	0.53	0.18	0.26	0.55
Concordance correlation coefficient	0.69	0.30	0.60	0.71
Bias correction factor	0.94	0.69	0.90	0.92
Modeling efficiency	0.52	0.13	0.42	0.55
Coefficient of model determination	1.85	4.22	2.25	1.79
Regression analysis				
Intercept (β ₀)				
Estimate	-0.08	0.41	-0.46	-0.12
SE	8.24	16.56	10.29	4.31
P-value (β ₀ = 0)	0.9973	0.9845	0.9608	0.9767
Slope (β ₁)				
Estimate	1.00	1.00	1.01	1.00
SE	0.12	0.33	0.16	0.12
P-value (β ₁ = 1)	0.9556	0.9989	0.9300	0.9332
MSEP source, % MSEP				
Mean bias	2.25	5.92	2.31	1.52
Systematic bias	0.01	0.01	0.01	0.01
Random error	97.7	94.1	97.7	98.5
Root MSEP				
Estimate	2.71	2.09	2.29	1.71
% of the mean	4.27	4.23	3.78	4.93

¹Obs: observed evaluation data set; MSEP: mean square error of the prediction. WH: withers height, BL: body length, BDL: body diagonal length, and RD: rib depth. r²: determination coefficient for the observed vs. predicted values.

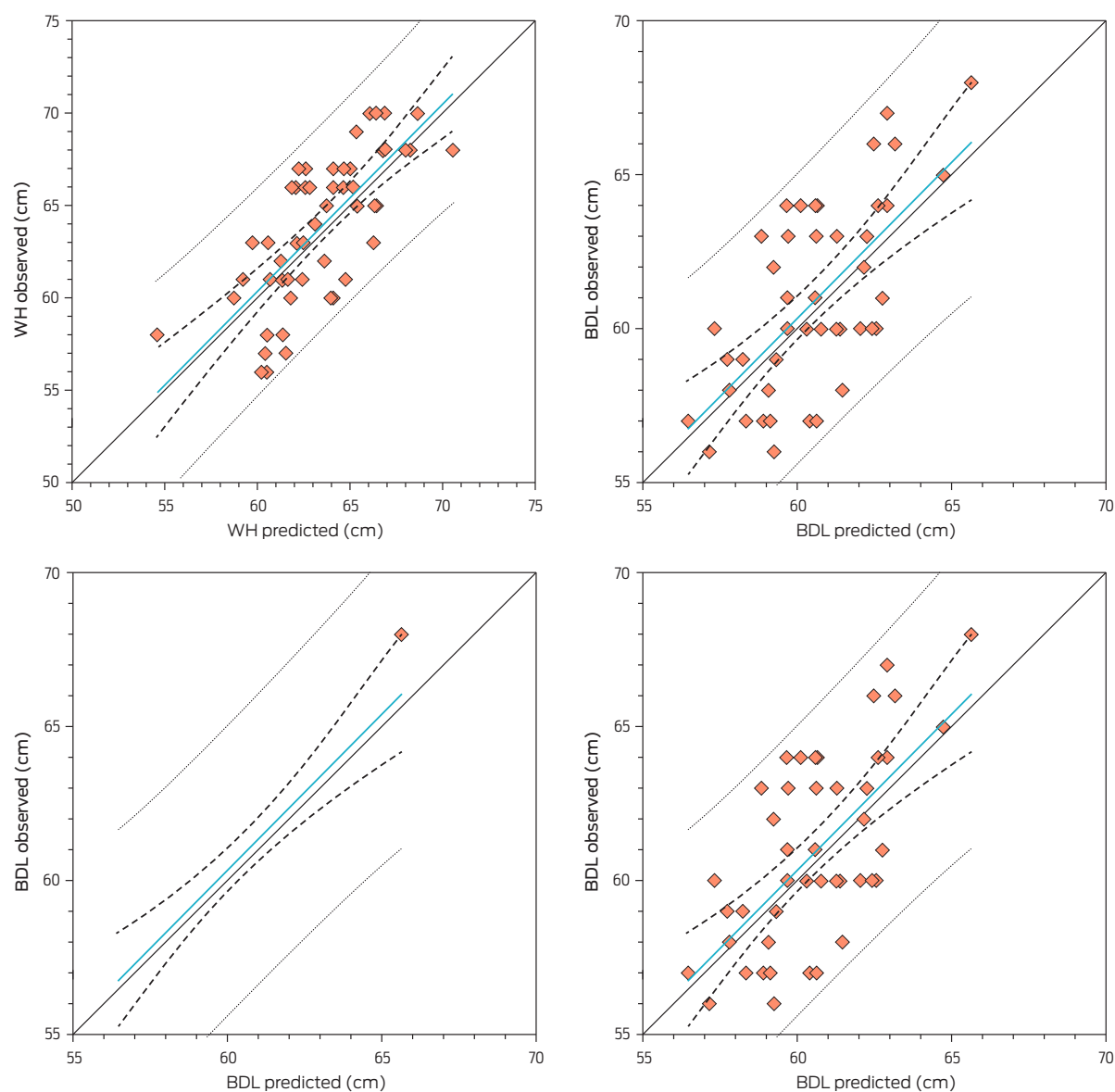


Figure 2. Relationship between the observed and predicted values for predicting biometric measurements of Pelibuey sheep using digital imaging. The solid line is $Y = X$, and the dotted line is the linear regression. WH: withers height, BL: body length, BDL: body diagonal length, and RD: rib depth. Data was based on 55 animals and one image per animal was used for DIA.

In addition, Morota et al.⁽²²⁾ mentioned that although the use of digital image analysis has great potential for BW estimation in livestock, some challenges still exist such as the automation of image data storage and statistical analysis. As a final point, although the results of the present study were not very promising, they delivered the basis for further developing the method in tropical sheep breeds. Moreover, it is necessary to evaluate the precision and accuracy of these equations using an independent dataset for verifying that they are not population-dependent. To improve equations accuracy, further studies should consider increasing the number of animals as well as several images obtained per animal. Another suggestion is to develop predictive equations for body weight using either the DIA approach or measuring tape.

Conclusions

Overall, the use of digital image analysis was able to predict biometric measurements in Pelibuey ewes with low to moderate precision ($r^2 > 0.18 \leq$ and $\leq r^2 0.55$) and accuracy (> 0.69). Factors that may affect the accuracy and precision of the relationships of biometric measurements *in vivo* to digital image analysis should be investigated. Future studies should address the relationships between digital image analysis and the prediction of body weight, body mass index, and body composition in animals from different physiological and production conditions. Also, to improve precision and accuracy, it is necessary to further evaluate that the equations are not population-dependent.

Data availability

The datasets generated during and/or analyzed during the current study are available from the corresponding author on reasonable request.

Acknowledgments

The authors are very grateful to Dr. José Manuel Piña Gutiérrez who provided the facilities of “El Rodeo” farm.

Funding statement

This study was partly sponsored by a research grant from the “Programa de Fomento a la Investigación” of Universidad Juárez Autónoma de Tabasco through the project “Eficiencia energética madre/cría en ovinos de pelo” [PFI: UJAT-DACA-2015-IA-02].

Conflicts of interest

Einar Vargas-Bello-Pérez, is member of the editorial board of Veterinaria México OA. Following the current journal's policies in this regard, he only participated as author during the editorial process of this submission. Moreover, all authors agreed to publish the peer review process along with the article.

Author contributions

Conceptualization: AJ Chay-Canul, R García-Herrera, E Vargas-Bello-Pérez.

Data curation: R García-Herrera, J Tapia-González, J Canul-Solís, F Casanova-Lugo, AT Piñeiro-Vázquez, R Portillo-Salgado.

Formal analysis: AJ Chay-Canul.

Funding acquisition: AJ Chay-Canul,

Investigation: AJ Chay-Canul, R García-Herrera, J Tapia-González, J Canul-Solís, F Casanova-Lugo, AT Piñeiro-Vázquez, R Portillo-Salgado, E Vargas-Bello-Pérez.

Methodology: AJ Chay-Canul, R García-Herrera, E Vargas-Bello-Pérez.

Project administration: AJ Chay-Canul.

Resources: AJ Chay-Canul

Software: AJ Chay-Canul, R García-Herrera.

Supervision: AJ Chay-Canul, R García-Herrera.

Validation: AJ Chay-Canul.

Visualization: AJ Chay-Canul, R García-Herrera, E Vargas-Bello-Pérez.

Writing-original draft: AJ Chay-Canul, R García-Herrera, E Vargas-Bello-Pérez.

Writing-review and editing: AJ Chay-Canul, R García-Herrera, J Tapia-González, J Canul-Solís, F Casanova-Lugo, AT Piñeiro-Vázquez, R Portillo-Salgado, E Vargas-Bello-Pérez.

References

1. Carabús A, Gispert M, Font-i-Furnols M. Imaging technologies to study the composition of live pigs: a review. *Spanish Journal of Agricultural Research*. 2016;14(3):e06R01. doi: 10.5424/sjar/2016143-8439.
2. Wu T, Gaw N, Xu Y, Li J, Wang L, Fu Y, Silva A, et al. Quantitative imaging system for cancer diagnosis and treatment planning: an interdisciplinary approach. *Informatics in Operations Research*. 2017;153–177. doi: 10.1287/educ.2017.0173.

3. Bertram CA, Klopfleisch R. The Pathologist 2.0: an update on digital pathology in Veterinary Medicine. *Veterinary Pathology*. 2017;54(5):756–766. doi: 10.1177/0300985817709888.
4. Argawal S, Chand S. Digital image forensic: a brief review. *Forensic Research & Criminology International Journal*. 2017;5(4):00161. doi: 10.15406/frcij.2017.05.00161.
5. Craigie CR, Navajas EA, Purchas RW, Maltin CA, Bünger L, Hoskin SO, Ross DW, Morris ST, Roehe R. A review of the development and use of video image analysis (VIA) for beef carcass evaluation as an alternative to the current EUROP system and other subjective systems. *Meat Science*. 2012;92(4):307–318. doi: 10.1016/j.meatsci.2012.05.028.
6. Cruz-Fernández M, Luque-Cobija MJ, Cervera ML, Morales-Rubio A, de la Guardia M. Smartphone determination of fat in cured meat products. *Microchemical Journal*. 2017;132:8–14. doi: 10.1016/j.microc.2016.12.020.
7. Nir O, Parmet Y, Werner D, Adin G, Halachmi I. 3D Computer-vision system for automatically estimating heifer height and body mass. *Biosystems Engineering*. 2018;173:4–10. doi: 10.1016/j.biosystemseng.2017.11.014.
8. Gomes RA, Monteiro GR, Assis GJF, Busato KC, Ladeira MM, Chizzotti ML. Technical note: Estimating body weight and body composition of beef cattle through digital image analysis. *Journal of Animal Science*. 2016;94(12):5414–5422. doi: 10.2527/jas.2016-0797.
9. McPhee MJ, Walmsley BJ, Skinner B, Littler B, Siddell JP, Cafe LM, Wilkins JF, Oddy VH, Alempijevic A. Live animal assessments of rump fat and muscle score in Angus cows and steers using 3-dimensional imaging. *Journal of Animal Science*. 2017;95(4):1847–1857. doi: 10.2527/jas.2016.1292.
10. Cominotte A, Fernandes AFA, Dorea JRR, Rosa GJM, Ladeira MM, van Cleef EHCB, Pereira GL, Baldassini WA, Machado Neto OR. Automated computer vision system to predict body weight and average daily gain in beef cattle during growing and finishing phases. *Livestock Science*. 2020;232:103904. doi: 10.1016/j.livsci.2019.103904.
11. Miller GA, Hyslop JJ, Barclay D, Edwards A, Thomson W, Duthie CA. Using 3D imaging and machine learning to predict liveweight and carcass characteristics of live finishing beef cattle. *Frontiers in Sustainable Food Systems*. 2019;3:30. doi: 10.3389/fsufs.2019.00030.
12. Martins BM, Mendes ALC, Silva LF, Moreira TR, Costa JHC, Rotta PP, Chizzotti ML, Marcondes MI. Estimating body weight, body condition score, and type traits in dairy cows using three dimensional cameras and manual body measurements. *Livestock Science*. 2020;236:104054. doi: 10.1016/j.livsci.2020.104054.
13. Liu D, He D, Norton T. Automatic estimation of dairy cattle body condition score from depth image using ensemble model. *Biosystems Engineering*. 2020;194:16–27. doi: 10.1016/j.biosystemseng.2020.03.011.
14. Weber VAM, Weber FL, Gomes RC, Junior ASO, Menezes GV, Abreu UGP, Belete NAS, Pistori H. Prediction of Girolando cattle weight by means of body measurements extracted from images. *Revista Brasileira de Zootecnia*. 2020;49:e20190110. doi: 10.37496/rbz4920190110.
15. Na Zhang AL, Pei Wu B, Xin Hua Jiang C, Chuan Zhong Xuan D, Yan Hua Ma E, Yong An Zhang F. Development and validation of a visual image analysis

- for monitoring the body size of sheep. *Journal of Applied Animal Research*. 2018;46(1):1004–1015. doi: 10.1080/09712119.2018.1450257.
16. Carabús A, Gispert M, Brun A, Rodríguez P, Font-i-Furnols M. *In vivo* computed tomography evaluation of the composition of the carcass and various cuts of growing pigs of three commercial crossbreeds. *Livestock Science*. 2014;170:181–192. doi: 10.1016/j.livsci.2014.10.005.
 17. Yan Q, Ding L, Wei H, Wang X, Jiang C, Degen A. Body weight estimation of yaks using body measurements from image analysis. *Measurement*. 2019;140:76–80. doi: 10.1016/j.measurement.2019.03.021.
 18. Khojastehkey M, Kalantar Neyestanaki M, Roudbari Z, Sadeghipanah H, Javaheri H, Aghashahi AR. Feasibility of body weight estimation of Kalkoohi camels using digital image processing. *Iranian Journal of Applied Animal Science*. 2020;10(2):333–340.
 19. Mollah BR, Hasan A, Salam A, Ali A. Digital image analysis to estimate the live weight of broiler. *Computers and Electronics in Agriculture*. 2010;72(1):48–52. doi: 10.1016/j.compag.2010.02.002.
 20. Mortensen AK, Lisouski P, Ahrendt P. Weight prediction of broiler chickens using 3D computer vision. *Computers and Electronics in Agriculture*. 2016;123:319–326. doi: 10.1016/j.compag.2016.03.011.
 21. Wishart H, Morgan-Davies C, Stott AW, Wilson A, Waterhouse T. Liveweight loss associated with handling and weighing of grazing sheep. *Small Ruminant Research*. 2017;153:163–170. doi: 10.1016/j.smallrumres.2017.06.013.
 22. Morota G, Ventura RV, Silva FF, Koyama M, Fernando SC. Big data analytics and precision animal agriculture symposium: machine learning and data mining advance predictive big data analysis in precision animal agriculture. *Journal of Animal Science*. 2018;96(4):1540–1550. doi: 10.1093/jas/sky014.
 23. Kongsro J. Estimation of pig weight using a Microsoft Kinect prototype imaging system. *Computers and Electronics in Agriculture*. 2014;109:32–35. doi: 10.1016/j.compag.2014.08.008.
 24. Chay-Canul AJ, García-Herrera RA, Salazar-Cuytún R, Ojeda-Robertos NF, Cruz-Hernández A, Fonseca MA, Canul-Solís JR. Development and evaluation of equations to predict body weight of Pelibuey ewes using heart girth. *Revista Mexicana de Ciencias Pecuarias*. 2019;10(3):767–777. doi: 10.22319/rmcp.v10i3.4911.
 25. Bautista-Díaz E, Salazar-Cuytun R, Chay-Canul AJ, García-Herrera RA, Piñeiro-Vázquez AT, Magaña-Monforte JG, Tedeschi LO, Cruz-Hernández A, Gómez-Vázquez A. Determination of carcass traits in Pelibuey ewes using biometric measurements. *Small Ruminant Research*. 2017;147:115–119. doi: 10.1016/j.smallrumres.2016.12.037.
 26. Canul-Solís J, Angeles-Hernandez JC, García-Herrera RA, del Razo-Rodríguez OE, Lee Rangel HA, Piñeiro-Vázquez AT, Casanova-Lugo F, Rosales-Nieto CA, Chay-Canul AJ. Estimation of body weight in hair ewes using an indirect measurement method. *Tropical Animal Health and Production*. 2020;52:2341–2347. doi: 10.1007/s11250-020-02232-7.
 27. O' Leary N, Leso L, Buckley F, Kenneally J, McSweeney D, Shalloo L. Validation of an automated body condition scoring system using 3D imaging. *Agriculture*. 2020;10(6):246. doi: 10.3390/agriculture10060246.

28. Ozkaya S, Bozkurt Y. The relationship of parameters of body measures and body weight by using digital image analysis in pre-slaughter cattle. *Archives Animal Breeding*. 2008;51(2):120–128. doi: 10.5194/aab-51-120-2008.
29. Tasdemir S, Urkmez A, Inal S. Determination of body measurements on the Holstein cows using digital image analysis and estimation of live weight with regression analysis. *Computers and Electronics in Agriculture*. 2011;76(2):189–197. doi: 10.1016/j.compag.2011.02.001.
30. Ozkaya S. Accuracy of body measurements using digital image analysis in female Holstein calves. *Animal Production Science*. 2012;52(10):917–920. doi: 10.1071/AN12006.
31. Chay-Canul AJ, Magaña-Monforte JG, Chizzotti ML, Piñeiro-Vázquez ÁT, Canul-Solís JR, Ayala-Burgos AJ, Ku-Vera JC, Tedeschi LO. Energy requirements of hair sheep in the tropical regions of Latin America. Review. *Revista Mexicana de Ciencias Pecuarias*. 2016;7(1):105–125.
32. AFRC. Energy and protein requirements of ruminants. Wallingford, UK: Agricultural and Food Research Council, CAB International; 1993. 159 pp.
33. Wongsriworaphon A, Arnonkijpanich B, Pathumnakul S. An approach based on digital image analysis to estimate the live weights of pigs in farm environments. *Computers and Electronics in Agriculture*. 2015;115:26–33. doi: 10.1016/j.compag.2015.05.004.
34. Tedeschi LO. Assessment of the adequacy of mathematical models. *Agricultural Systems*. 2006;89(2-3):225–247. doi: 10.1016/j.agsy.2005.11.004.
35. Cochran WG, Cox GM. *Experimental Design*. New York, US: John Wiley and Sons; 1957. 615 pp.
36. Loague K, Green RE. Statistical and graphical methods for evaluating solute transport models: overview and application. *Journal of Contaminant Hydrology*. 1991;7(1-2):51–73. doi: 10.1016/0169-7722(91)90038-3.
37. Mayer DG, Butler DG. Statistical validation. *Ecological Modelling*. 1993;68(1-2):21–32. doi: 10.1016/0304-3800(93)90105-2.
38. Lin LI. A concordance correlation coefficient to evaluate reproducibility. *Biometrics*. 1989;45(1):255–268. doi: 10.2307/2532051.
39. Wang Y, Yang W, Winter P, Walker L. Walk-through weighing of pigs using machine vision and an artificial neural network. *Biosystems Engineering*. 2008;100(1):117–125. doi: 10.1016/j.biosystemseng.2007.08.008.
40. Lasfeto DB, DaudLetik M. A measuring weight model of Timor's beef cattle based on image. *International Journal of Engineering and Technology*. 2017;9(2):677–688. doi: 10.21817/ijet/2017/v9i2/170902089.